

more capabilities.
new possibilities.



**A Systematic Approach for
Developing High Quality
LC/MS/MS Methods for Regulated
Bioanalysis**

Patrick Bennett
Pharma Marketing Director

Seminar Topics

- The realities and challenges of fast method development
- Means to develop high quality method
- Case studies
- Conclusion and Summary

Traditional Approach for Method Development

- Relies on personal experience and preference of the method development (MD) scientist
- Produce personalized method
- Conduct MD experiment linear “trail and error”
- Require longer MD time
- Result in less robust method
- Difficult to transfer
- Generate large inventory supplies and reagents

Need to change and standardize MD process!

Why Standardize Method Development Process

- Fewer variables – supplies, reagents, chemicals
- Better learning, common language, faster training
- Improve documentation
- Enhance MD team's technical expertise
- Simplify MD experiments
- Require less experienced scientific staff

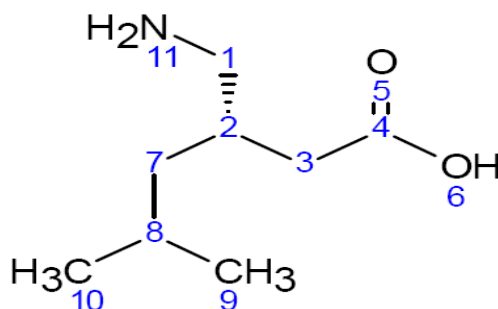
Concepts to Develop High Quality GLP Method

- Apply tighter acceptance criteria for MD data
- Understand project scope and molecule
- Standardize method development experiment
- Pre-emptive testing
- Manage phospholipids
- Evaluate linearity and carryover
- Monitor IS area response (Tandem Labs Presentation)
- Documentation and compliance

Tighter Acceptance Criteria for MD Data

- Expect better data from MD scientist who knows the assay the best
- Raises the confidence level in the assay
- More accurate stability data
- Proposed criteria:
 - Std /QC Precision/accuracy: $\leq \pm 10\%$ ($\leq \pm 15\%$ LLOQ)
 - Selectivity: 10 lots : $\leq \pm 10\%$ ($\leq 15\%$ LLOQ)

Case Study 1: Know Your Molecule: Pregabalin (zwitterionic)



Single-valued Properties

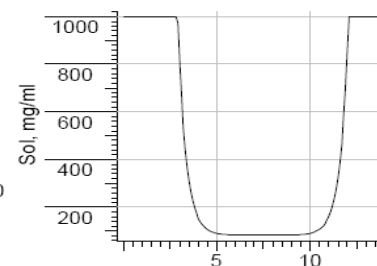
Name	Value	Error
LogP	1.12	0.23
MW	159.23	-
PSA	63.32	-
FRB	6	-
HDonors	3	-
HAacceptors	3	-
Rule Of 5	0	-
Molar Refractivity, cm ³	44.13	0.3
Molar Volume, cm ³	159.65	3
Parachor, cm ³	396.03	4
Index of Refraction	1.46	0.02
Surface Tension, dyne/cm	37.87	3
Density, g/cm ³	1	0.06
Polarizability, 10e-24 cm ³	17.49	0.5
Boiling Point, °C	274.03	23

pKa Results

Acidic/Basic	Acidic/Basic	Apparent pKa Value	Error
11	MB	11.31	0.1
6	MA	4.23	0.1

Solubility Results

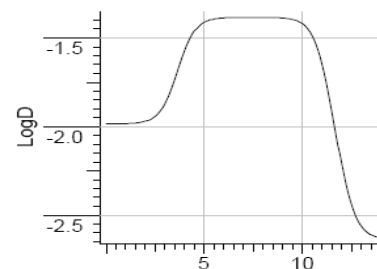
pH	Sol, mg/ml	Flags	%
1	1000	B	100
2	1000	B	99
5.5	84.87	Z	97
6.5	82.78	Z	100
7.4	82.6	Z	100
10	90.25	AZ	9 / 91
12	847.59	AZ	90 / 10



Intrinsic Solubility, mg/ml	20.6921
Intrinsic Solubility, log(S, mol/l)	-0.8862
Solubility in Pure Water at pH = 7.49, mg/ml	82.5956

LogD Results

pH	LogD
1	-1.98
2	-1.97
5.5	-1.39
6.5	-1.38
7.4	-1.38
10	-1.42
12	-2.22



Standardization of Sample Preparation Screening

- Use protocols for each experiment
- Build a common repository of reagents, SPE/SLE plates and LC columns.
- Conduct multiple experiments in parallel
- Treat all compounds as unknown

Case study 2: Fast MD for Guanfacine and Raltegravir

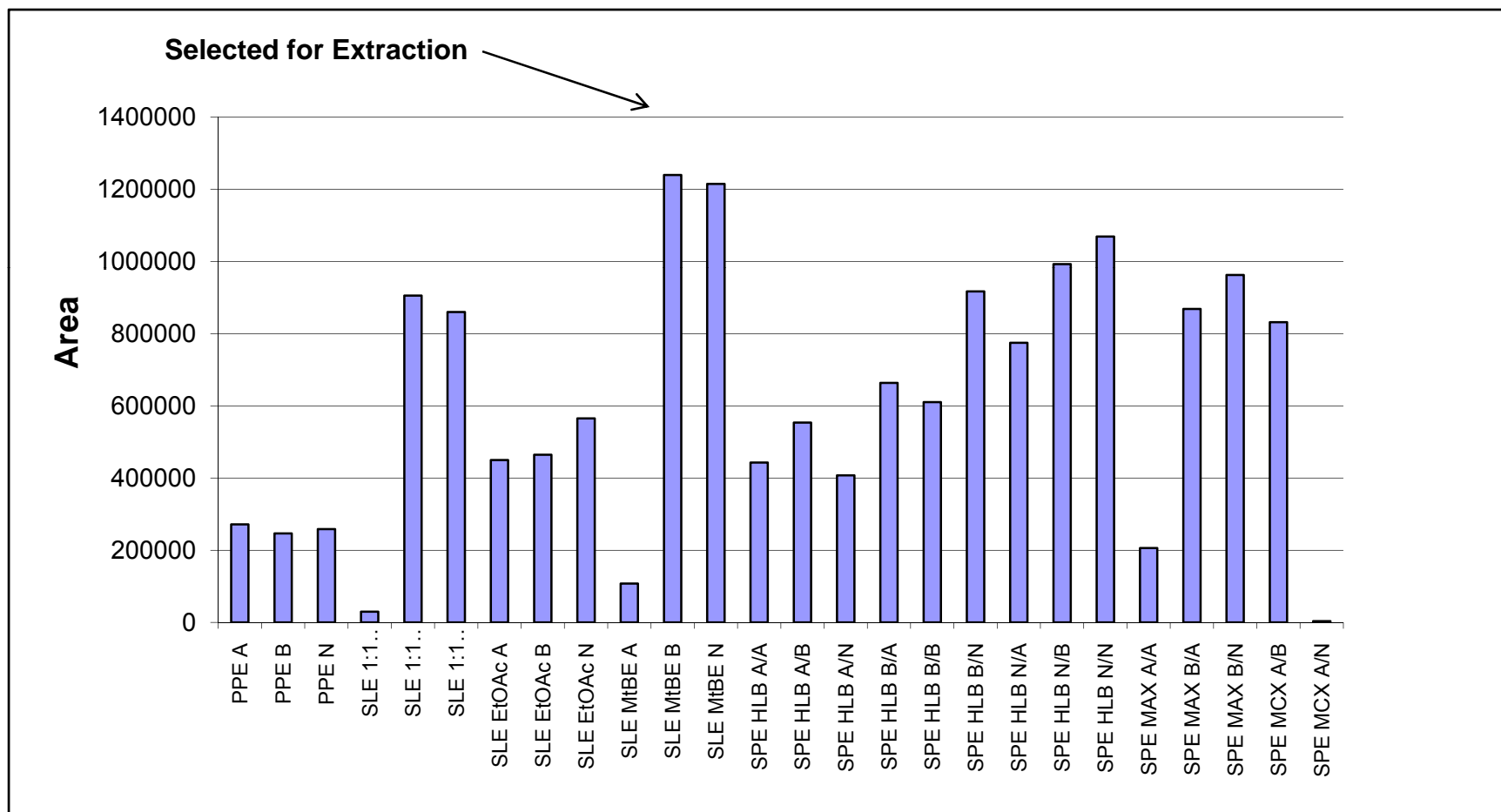
- Goal: Develop separate LC/MS/MS assays for Guanfacine and Raltegravir
- Approach: Conduct sample screening experiments together
- Time to complete: two days

Case study 2: Fast MD for Guanfacine and Raltegravir

- Day 1: sample preparation screening protocol
- Experiment design:
 - PPE: three pHs
 - SPE: Oasis MD plate combined with A, B, N washing and elution
 - SLE: three pHs, three organic solvents
- Total samples: 78

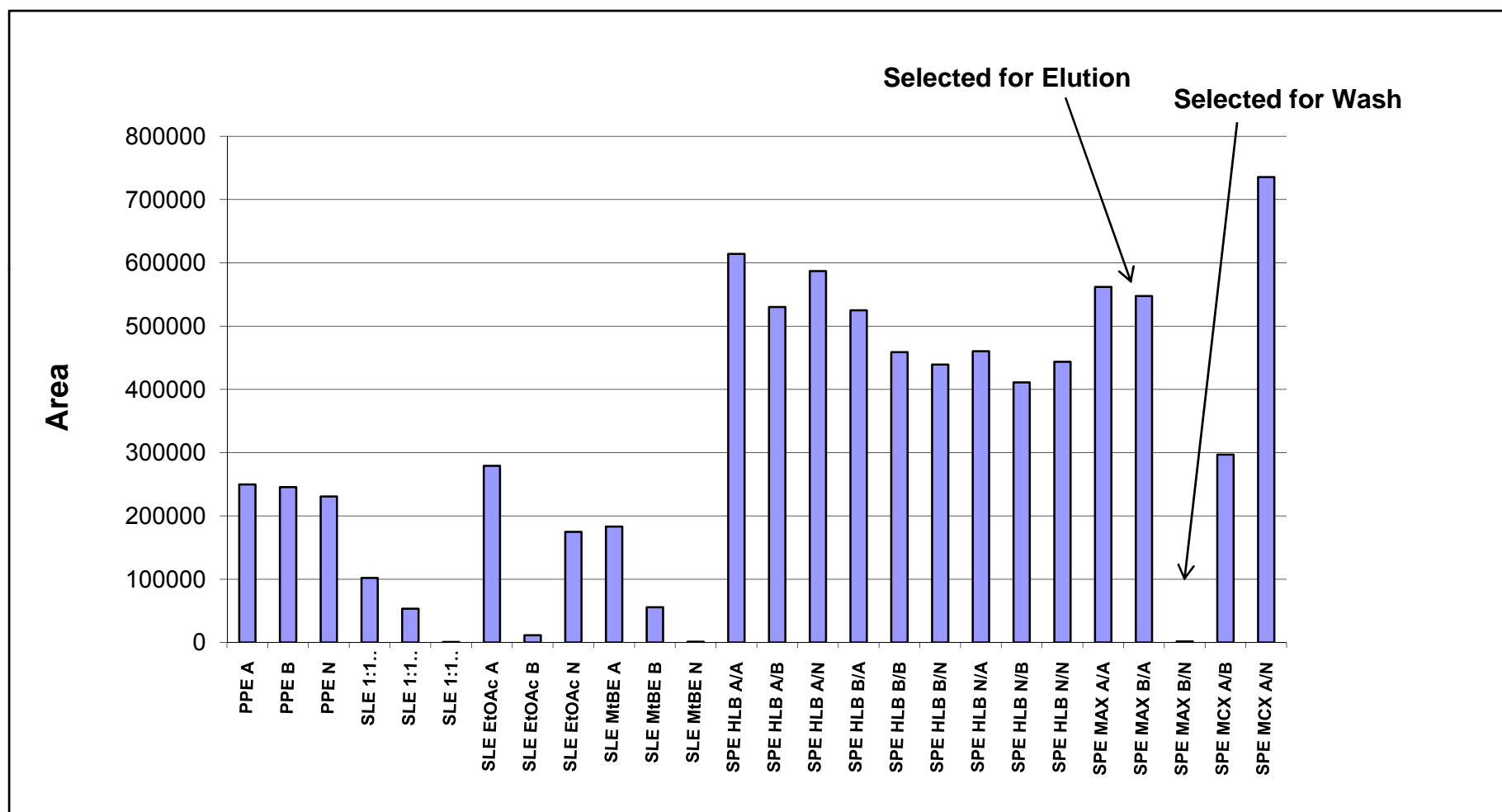
Case study 2: Fast MD for Guanfacine and Raltegravir

Guanfacine



Case study: Fast MD for Guanfacine and Raltegravir

Raltegravir

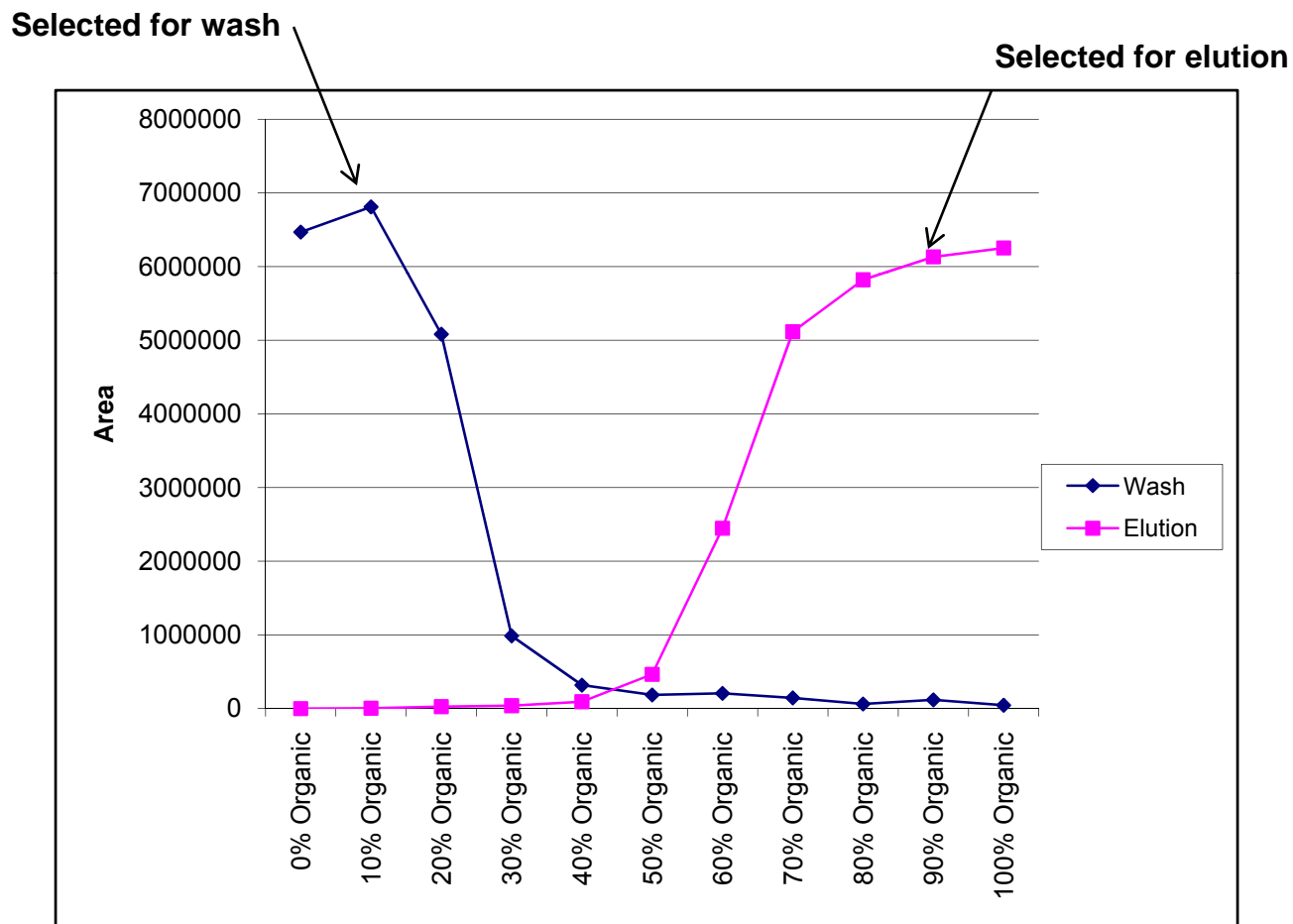


Case study 2: Fast MD for Guanfacine and Raltegravir

- Day 2: SPE wash/elution optimization for Raltegravir
- Experiment design:
 - Test MAX plate for B/A condition
 - Screen 10% organic component increments
- Bracket optimum wash/elution conditions
 - Use strongest wash that doesn't elute analyte
 - Use weakest elution that completely elutes analyte
- Total samples: 44

Case study 2–Fast MD for Guanfacine and Raltegravir

Raltegravir

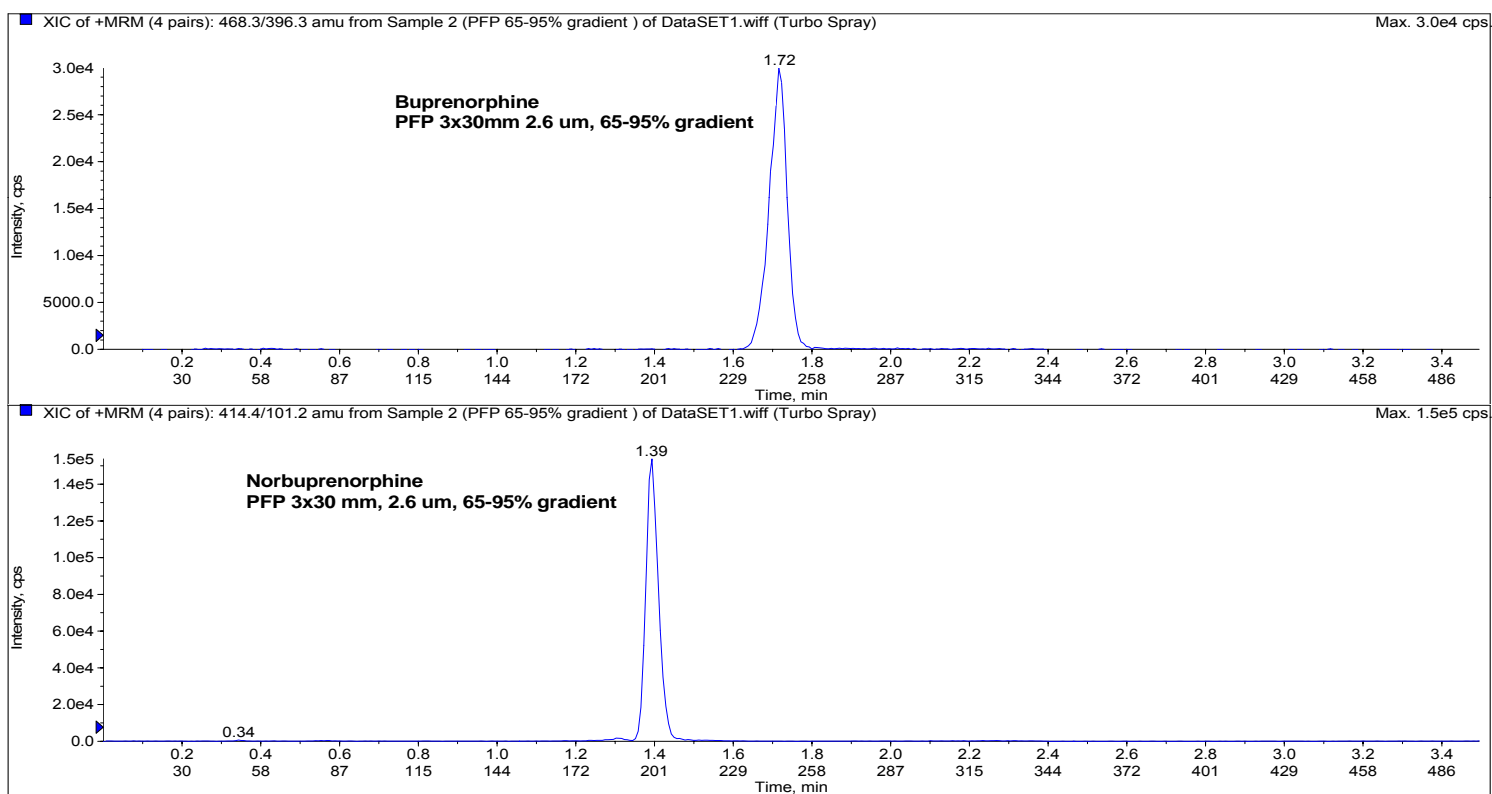


Standardization of LC/MS/MS development

- Standardize LC/AS cart
 - Add switching valve-diversion, backflush, trapping
- MS Tuning:
 - Tuning analyte under APCI/ESI, Neg/pos
 - Identify at least 2-3 SRM transitions
- LC column selection:
 - C18, phenyl-hexyl, Phenyl, HILIC, PFP (Pentafluorophenyl) column
 - HPLC vs. UHPLC

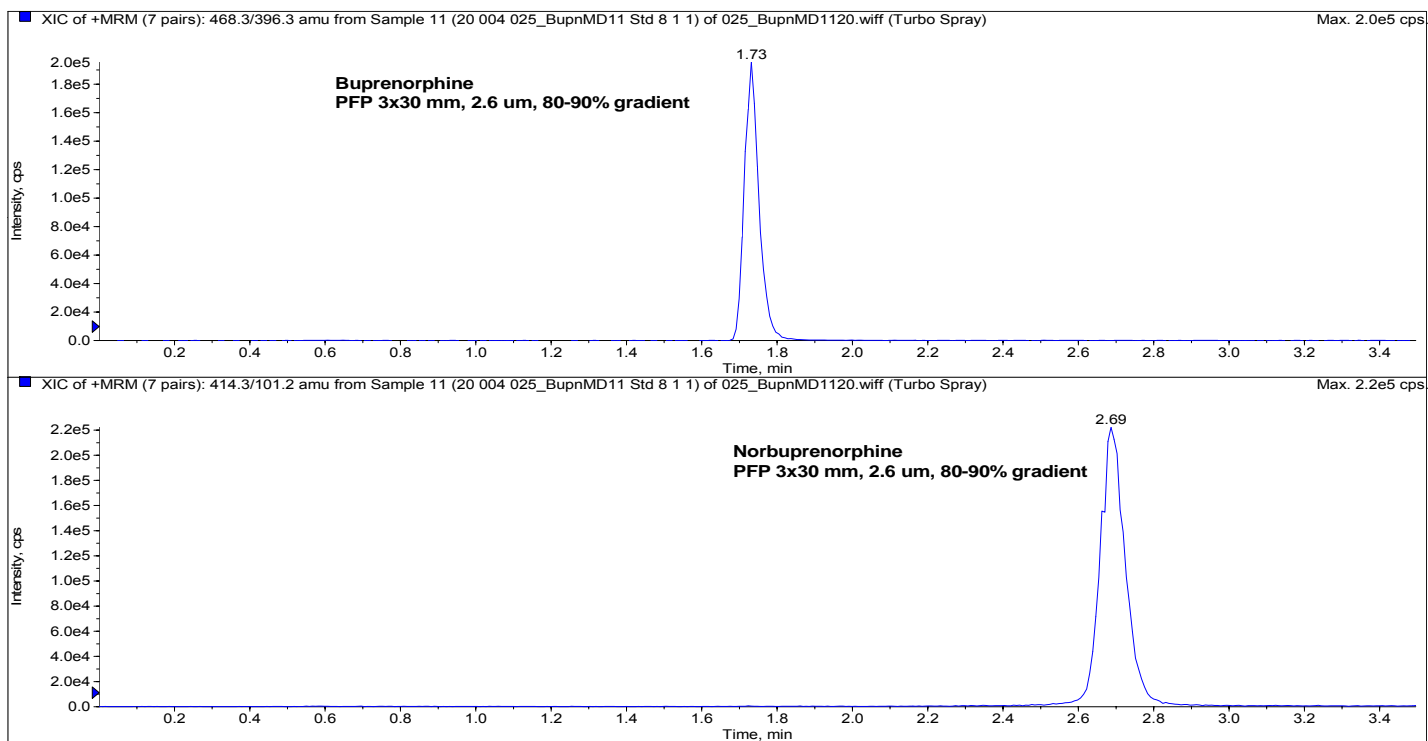
Case study 3 – PFP: Reversed vs. Normal Phase Mechanism

65-95% gradient – reversed phase chromatography



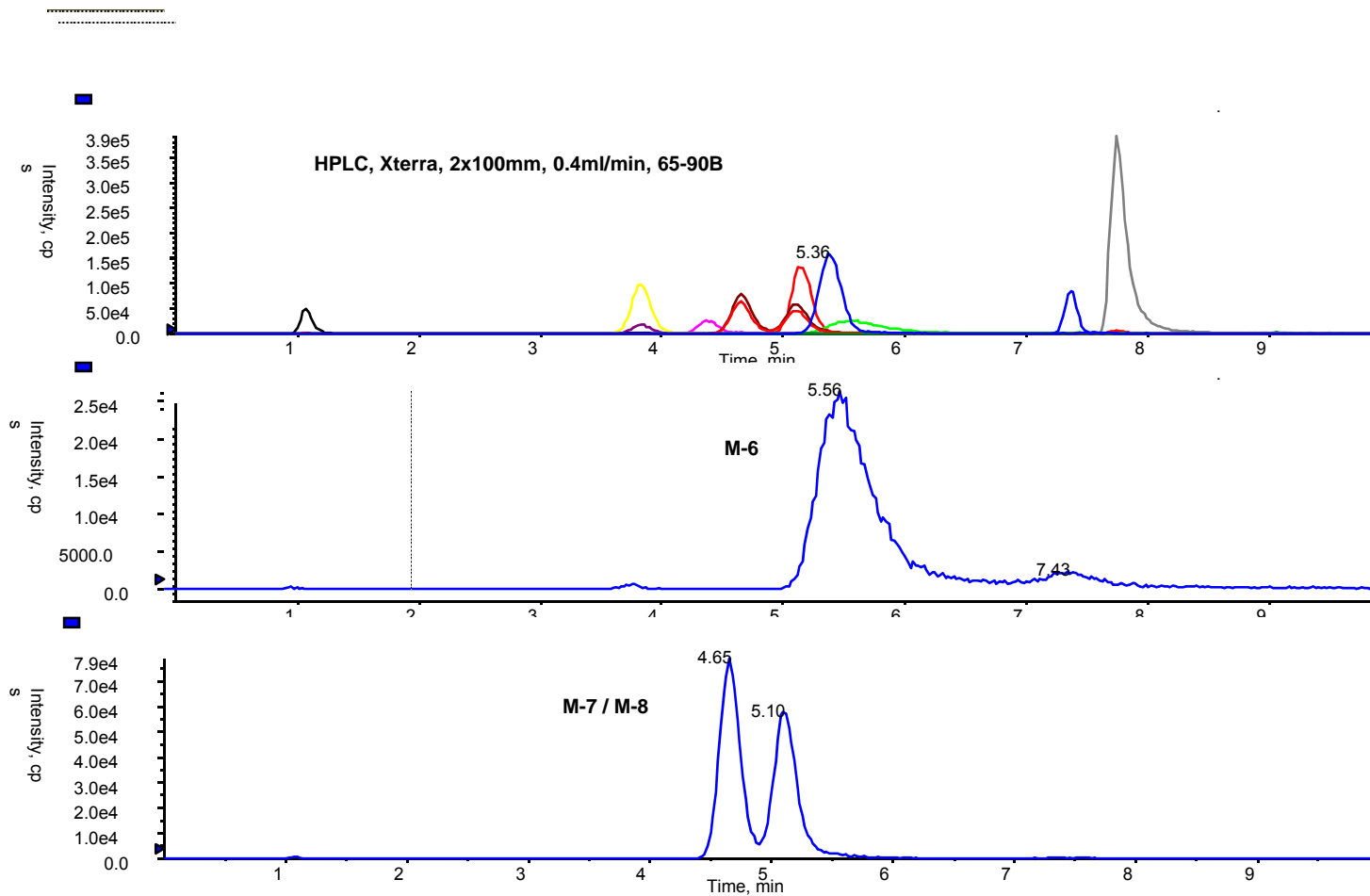
Case study 3 – PFP column: Reversed vs. normal phase

80-90% gradient – normal phase chromatography



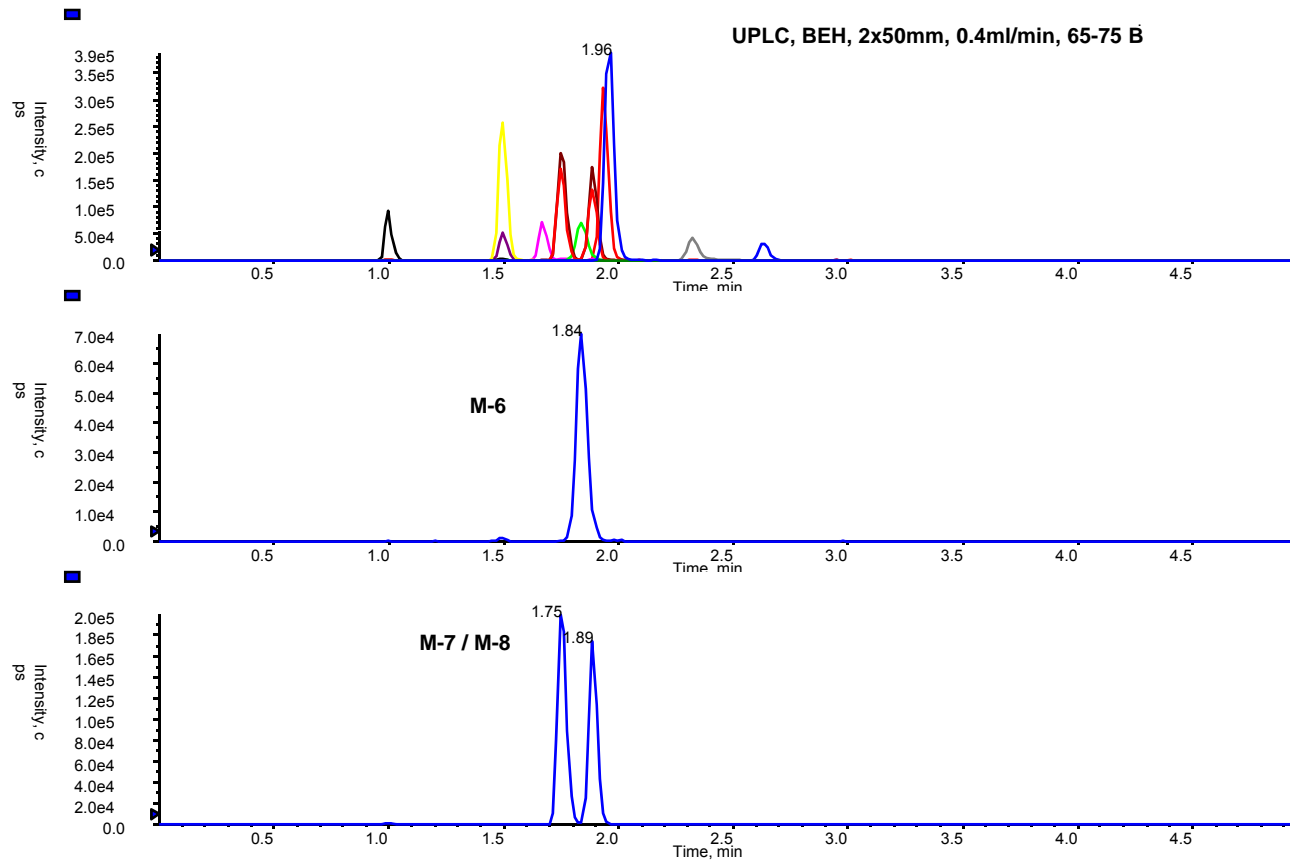
Case study 4: HPLC vs. UHPLC-Improving Resolution and Efficiency

HPLC, 10 minutes



Case study 4: HPLC vs. UHPLC-Improving Resolution and Efficiency

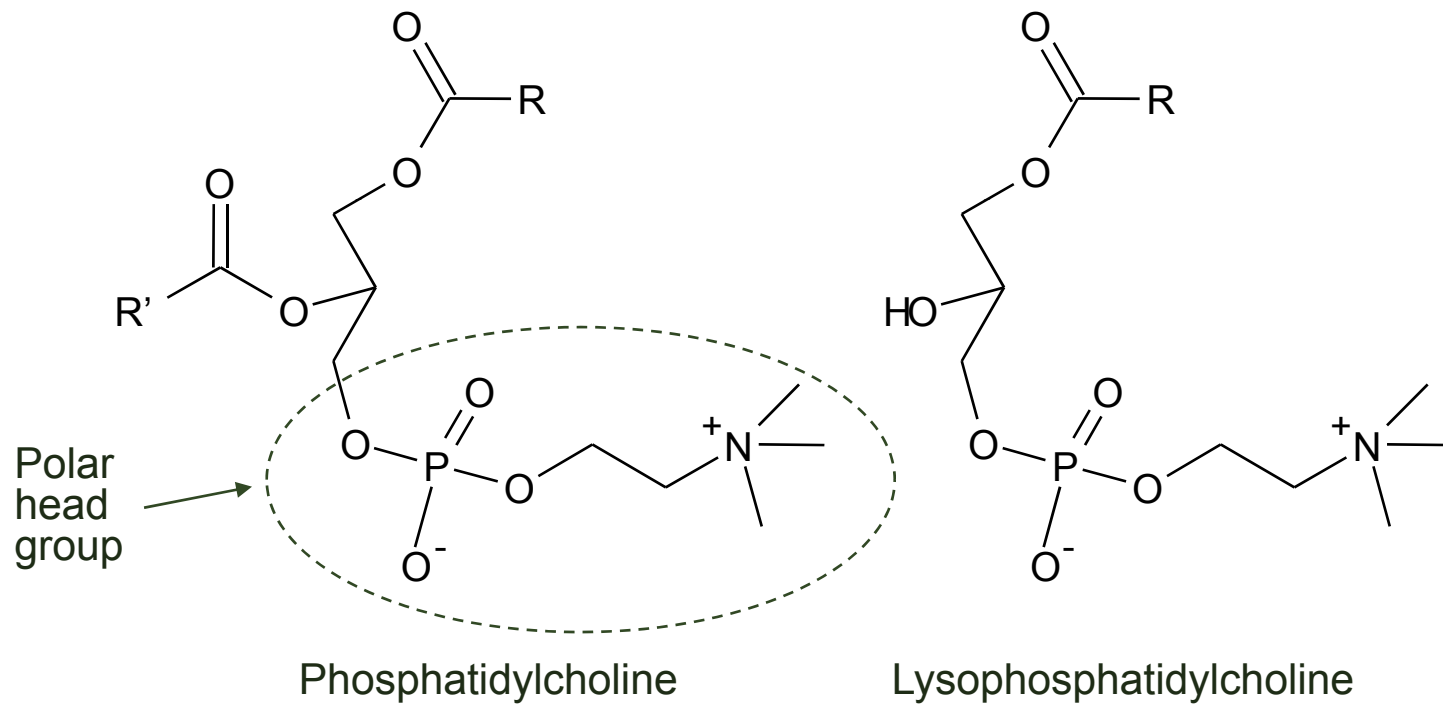
UHPLC, 4.5 minutes



Known Matrix Effectors

- Salt (problem in urine; L/L, SPE and turbulent flow remove)
- Proteins, peptides (most removed by all techniques)
- **Phospholipids - “The Biggest Problem”**
- Others
 - Other lipids (e.g., fatty acids – negative mode effects)
 - Dosing formulations
 - Unknowns...

Phospholipids



$R, R' = C_{12} - C_{18}$

Phospholipids Characteristics

- Ubiquitous (present in all species)
- High-levels (5-20 mg/ml in mammalian plasma)
- Chromatographically disperse; overall, relatively late-eluters in RP
- Accumulate on/bleed off LC column
- Surfactant behaviors— significant “matrix effectors”, especially in ESI/MS
- Matrix-variable (e.g., human diets)
- Unstable – degrade to fatty acids and head groups (effect on incurred sample repeat analyses)

Managing phospholipids

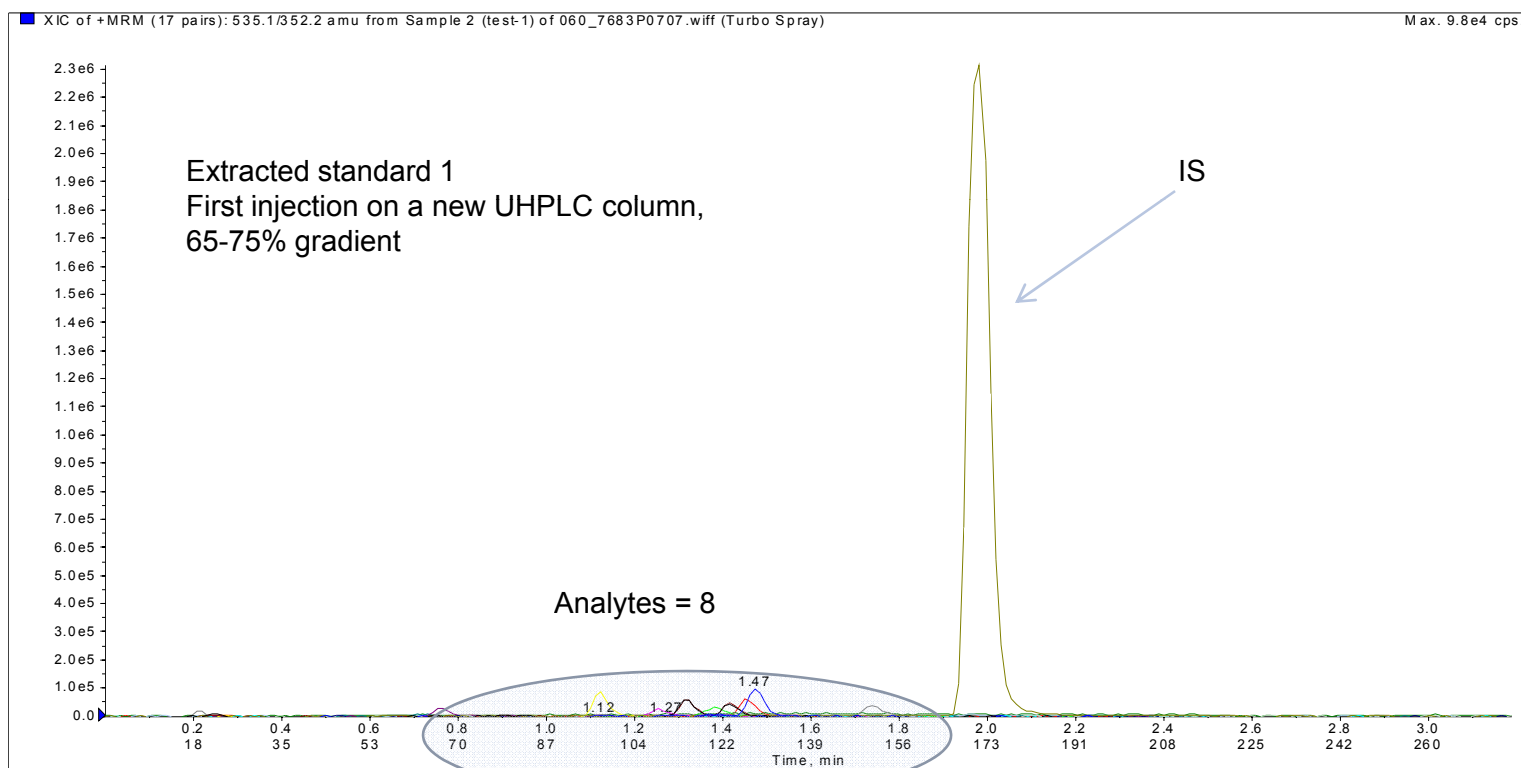
- Monitor major PL transitions all the time:
 - 496, 524 → 184 (LysoPC)
 - 704, 758, 806 → 184 (PC)
- Standardize LC/AS cart with two switching valves
- Decide if need to set up:
 - Guard column trapping
 - Column backflush
 - Column forward flush
 - Diversion

Case Study 5: Elution of phospholipids-forward flush

- Proprietary assay in human plasma (eight analytes and one IS)
- Protein precipitation sample preparation
- Issues: PL bleed
- Solution: LC column forward flush to remove phospholipids

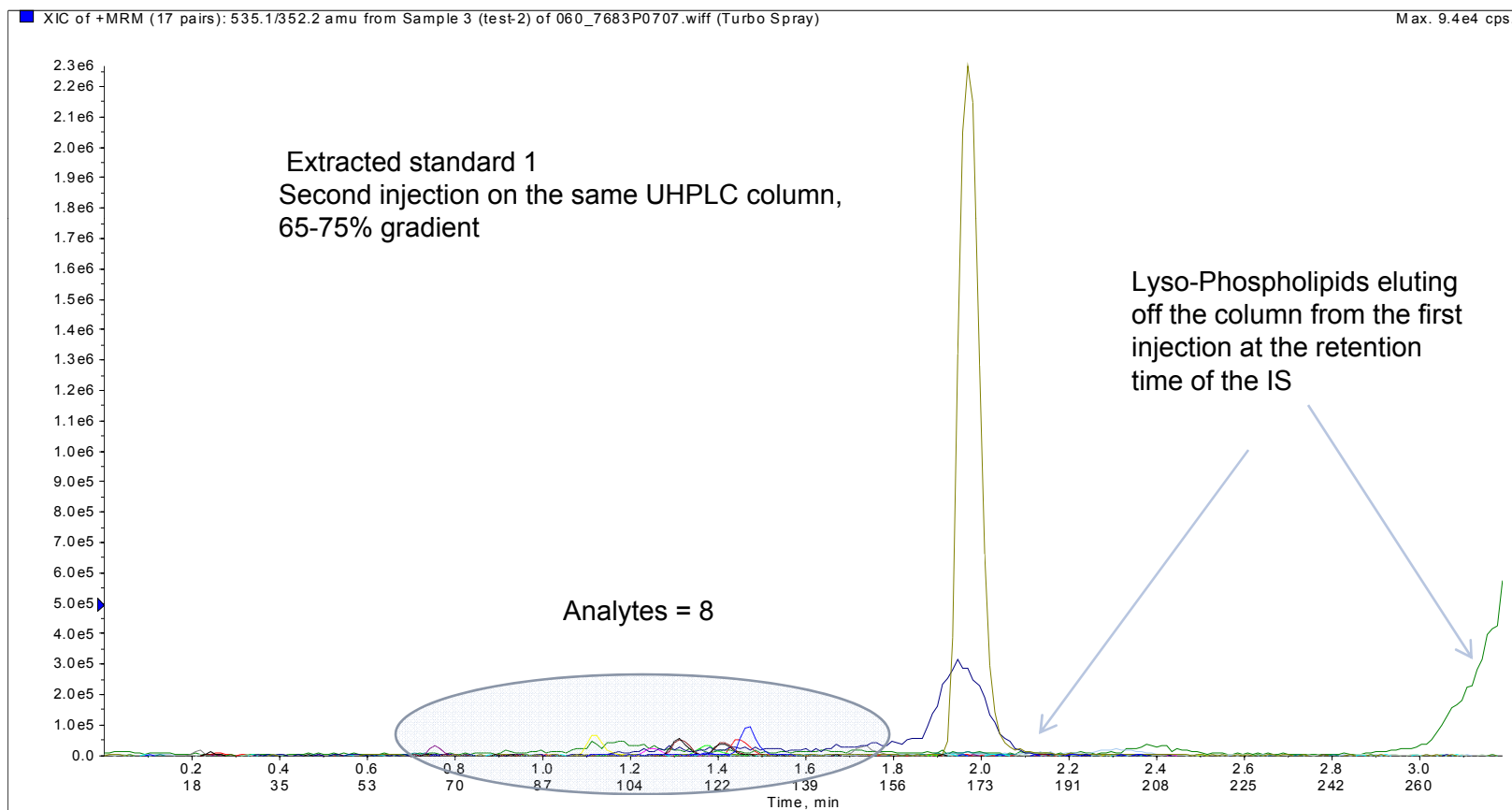
Case Study 5: Elution of phospholipids-forward flush

First injection on a brand new UHPLC column (65-75%)



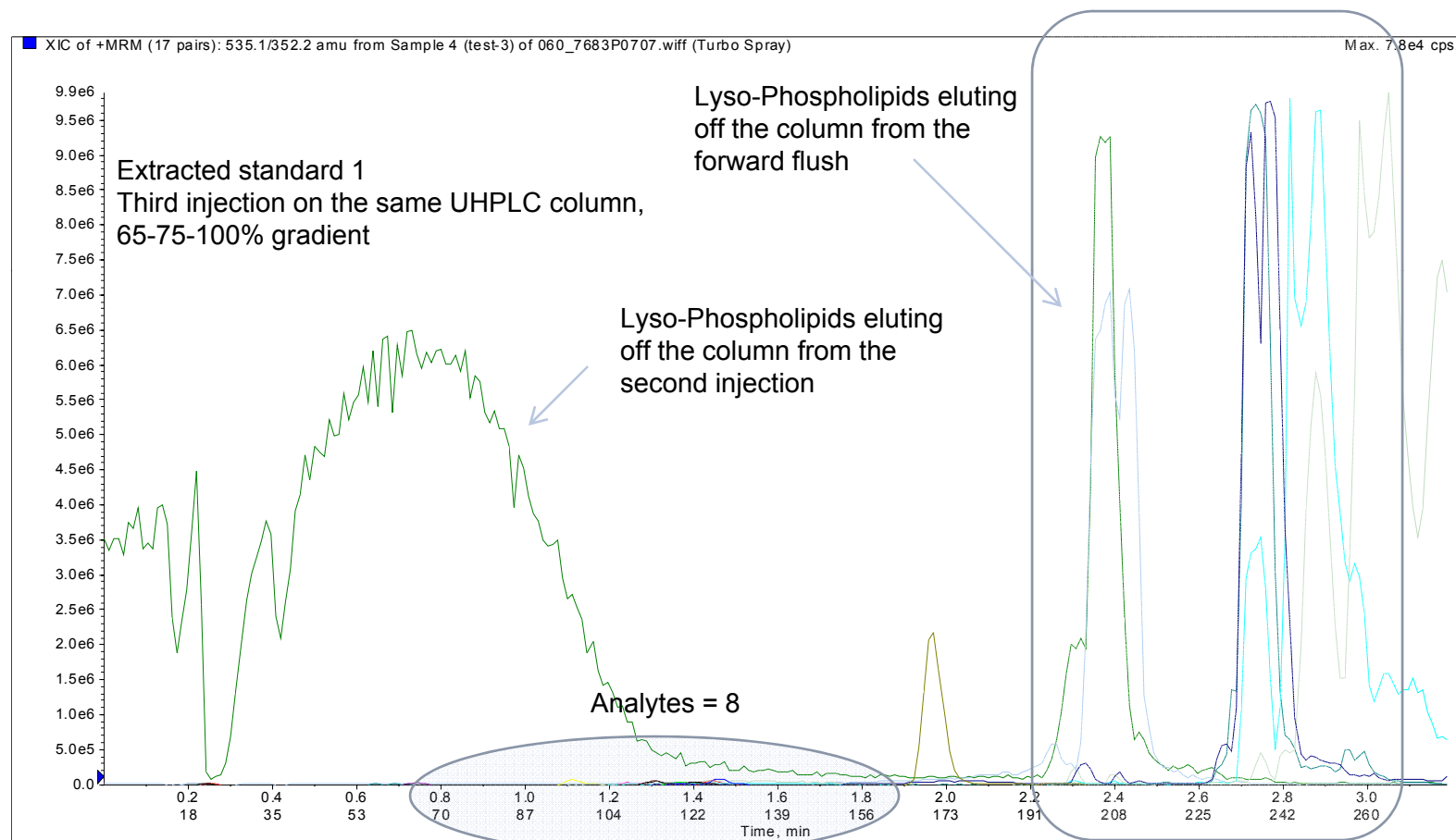
Case Study 5: Elution of phospholipids-forward flush

Second injection on the same column (65-75%)



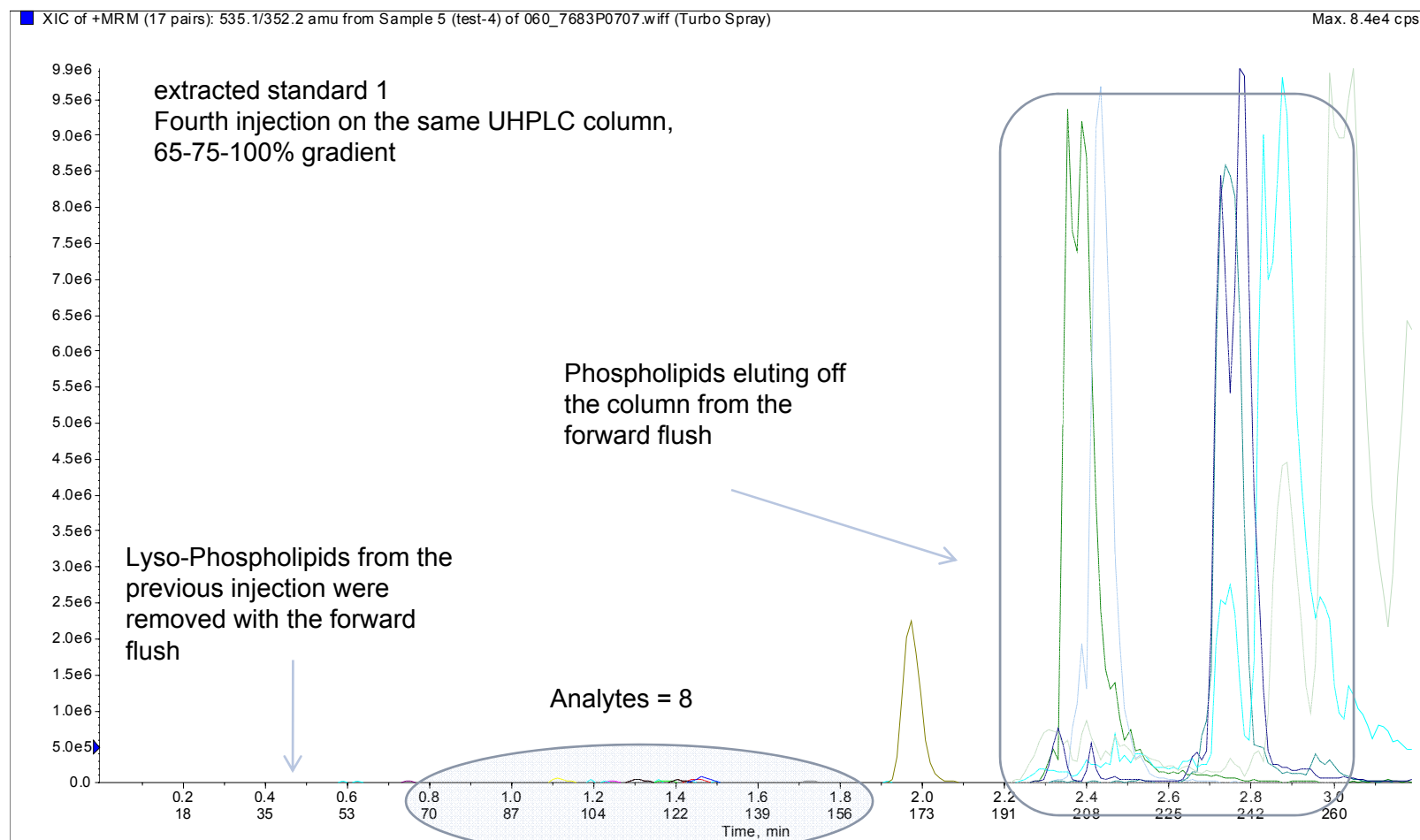
Case Study 5: Elution of phospholipids-forward flush

Third injection on the same column (65-75-100%)



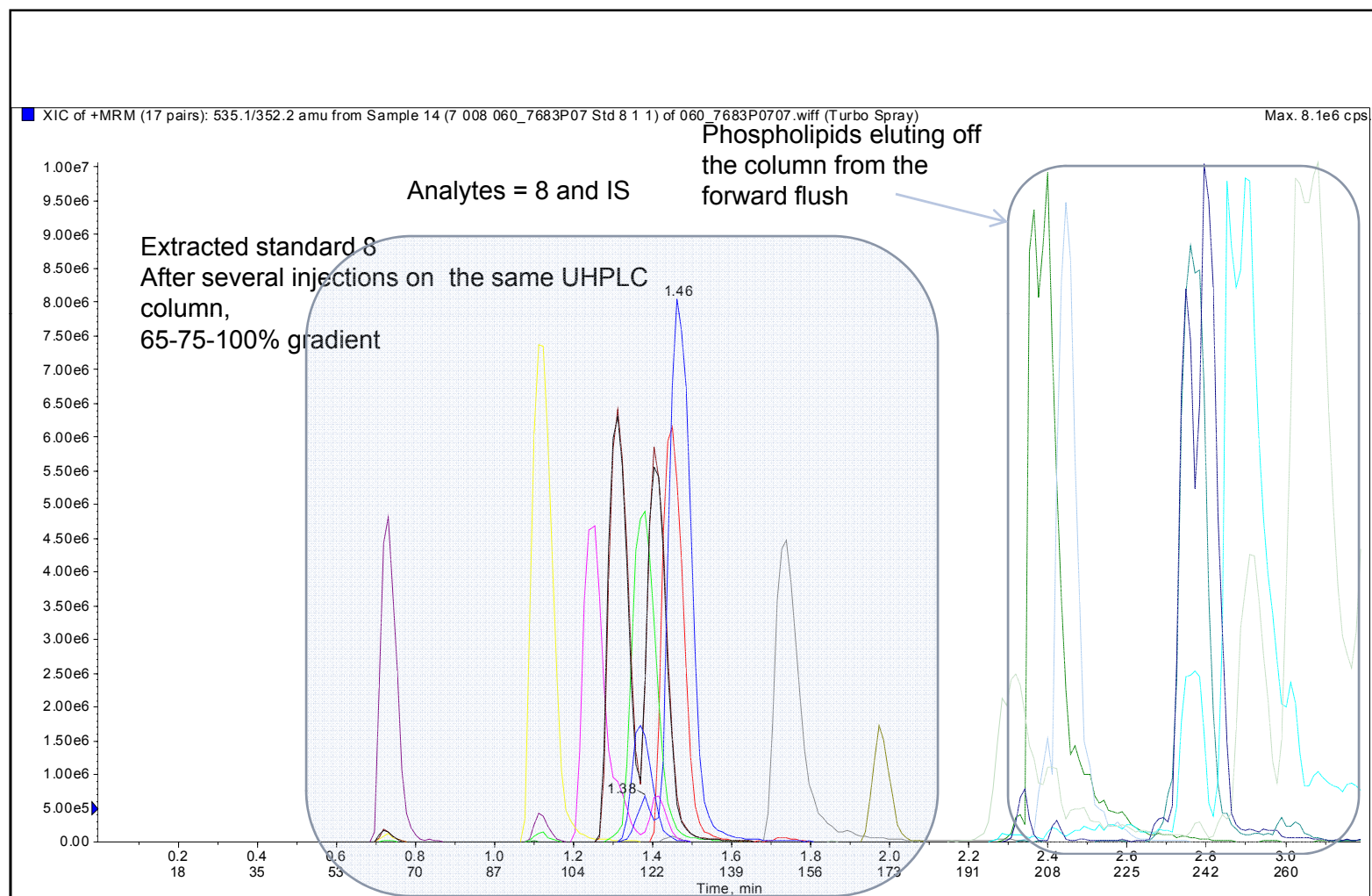
Case Study 5: Elution of phospholipids-forward flush

Fourth injection on the same column (65-75-100%)



Case Study 5: Elution of phospholipids-forward flush

Final condition, High Std, 65-75-100%



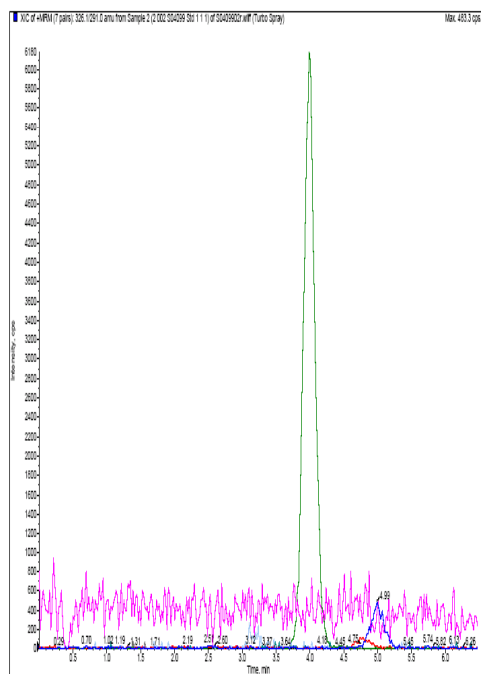
Case study 6: Splitting curve and increased PL background

- Midazolam and hydroxymidazolam in human plasma
- Protein precipitation sample preparation
- Issues: Splitting curve and raised PL background
- Solution: LC column backflush to remove phospholipids (10:90 water/acetone)

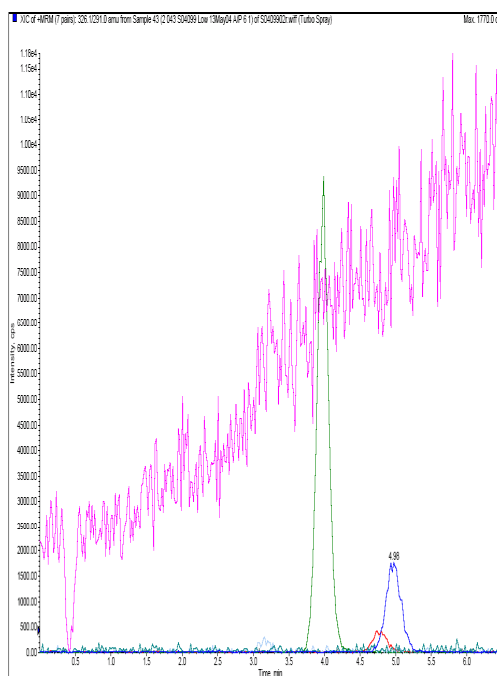
Case study 6-Splitting Curve and Raised PL Background

No column backflush

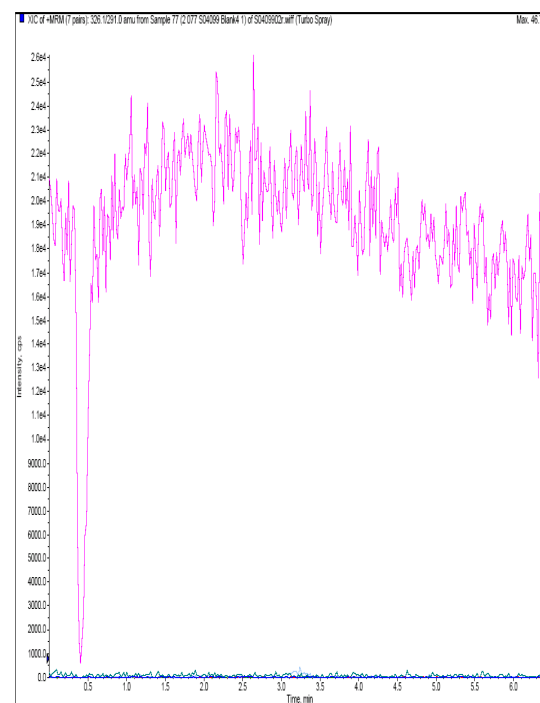
Sample #1



Sample #47

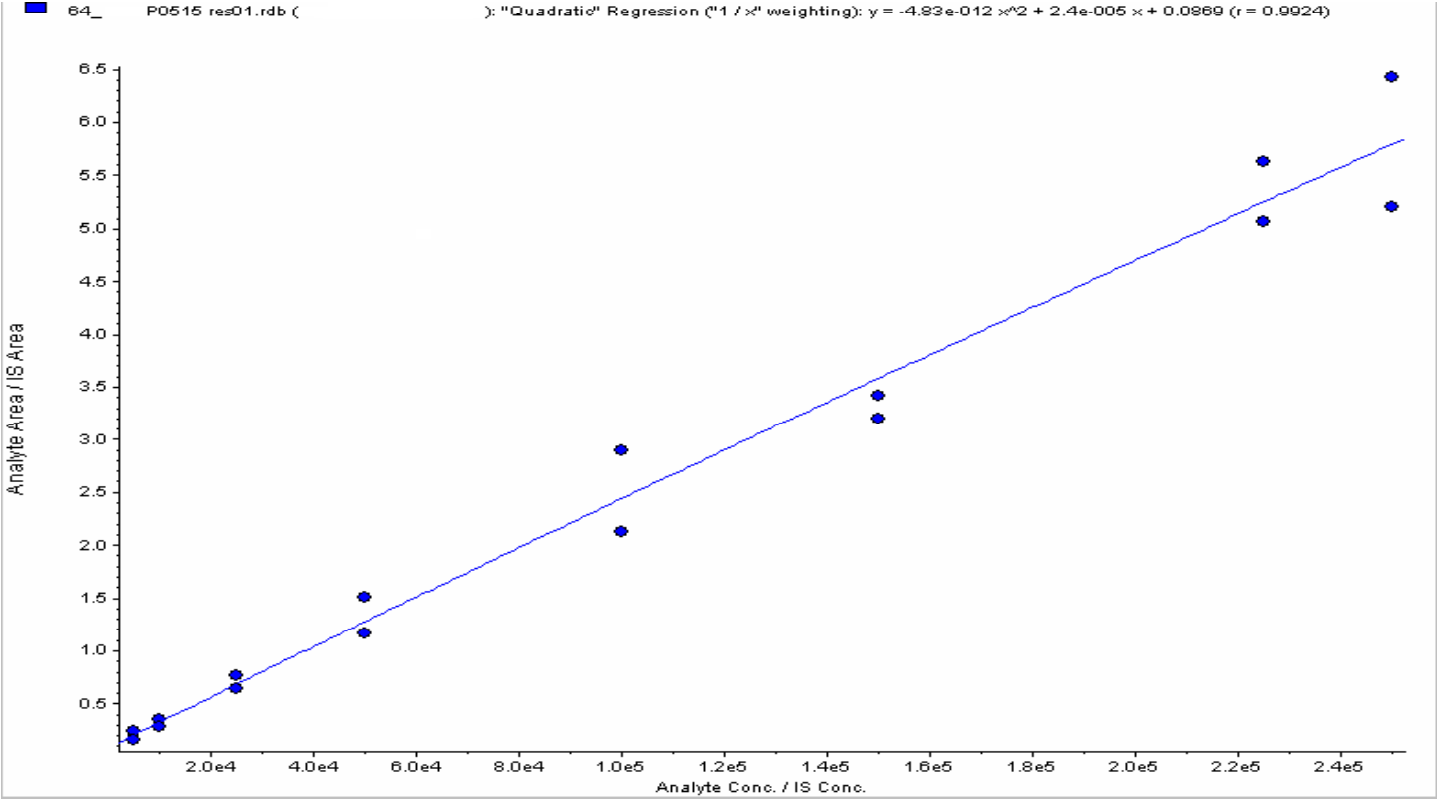


Sample #77



Case study 6-Splitting Curve and Raised PL Background

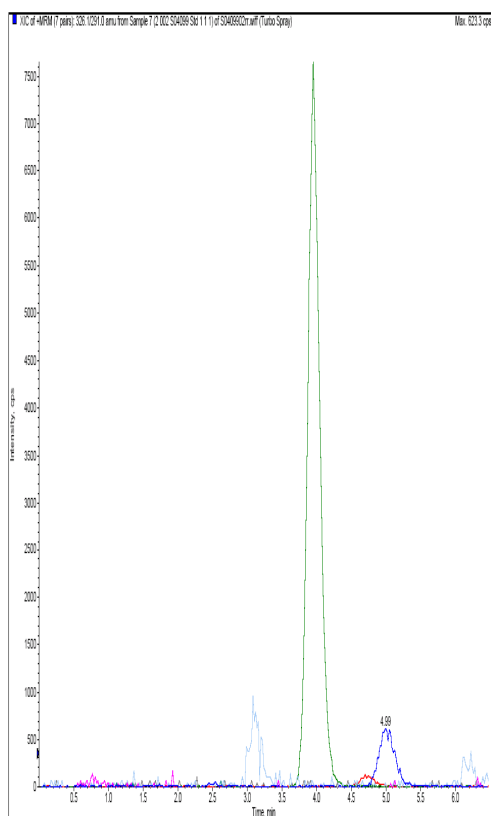
No column backflush



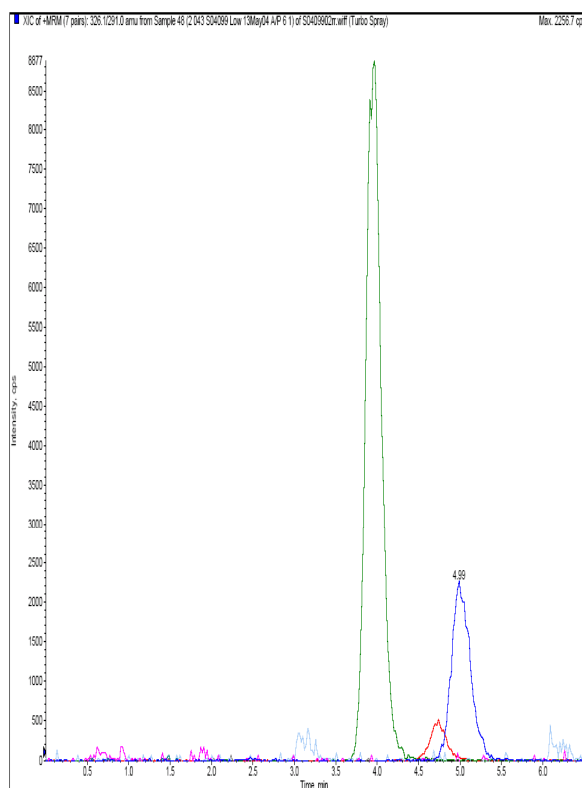
Case study 6-Splitting Curve and Raised PL Background

With column backflush

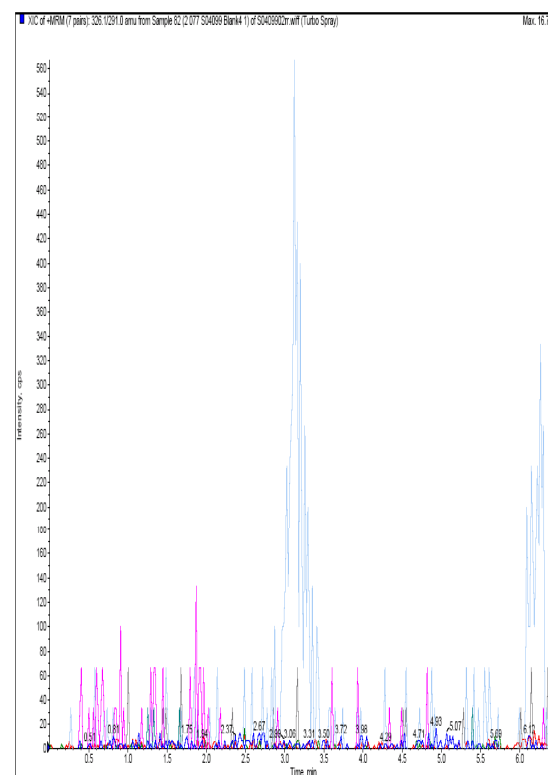
Sample #1



Sample #44

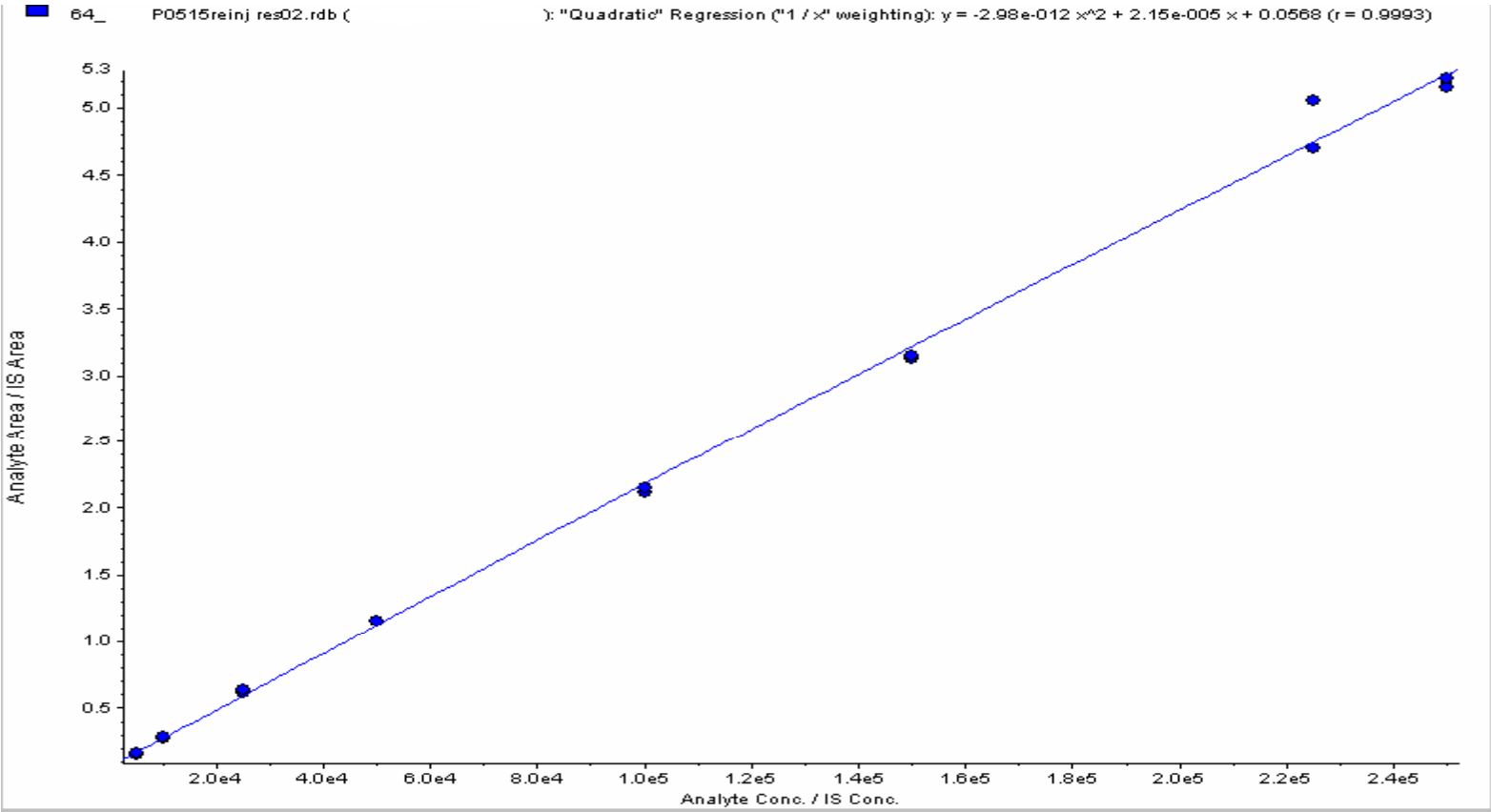


Sample #77



Case study 5-Splitting Curve and Raised PL Background

With column backflush



Evaluate linearity, carryover and stress test

Linearity:

- APCI is preferred over ESI
- Narrow linear range if possible

Carryover:

- Use isocratic or shallow gradient if possible
- Identify if injector Vs. LC column carryover

Stress test:

- Evaluate linear range at 2x below target LLOQ and 2x above ULOQ
- Inject test batch consecutively at 5 μ L, 10 μ L, 20 μ L or 30 μ L

Documentation and Compliance

- Regulated bioanalysis involves in multiple personnel and department.
- It is important to capture of all information for each critical stage of process.
- Utilization of three-tiered checklists:
 - Pre-method development (Pre-MD) checklist
 - Pre-method validation (Pre-MV) checklist
 - Post-method validation (Post-MV) checklist

Pre-MD Checklist

Gather information:

- Species, anticoagulant, aliquots, target LLOQ
- co-admin drugs, reference material info, ISTD info, metabolite info
- calibration range, previous methodology, stability, stickiness info
- If structure is available, use software or others to chemical-physical property
- Pay attention to specific functional groups

Pre-MV Checklist-Part 1

MD scientist self checklist

- Preemptive experiments
- Qualification batch with validation QCs
- Selectivity evaluation with 10 lots
- Carryover and linearity assessment
- Recovery evaluation
- Interference evaluation (analyte only and/or co-administered drug)
- Consecutive reinjection
- Phospholipid profile assessment

Pre-MV Checklist-Part 2

MV Qualification data

- Chromatogram prints:
 - LLOQ (S/N should be $> 10:1$)
 - Carryover Blank (contribution $< 20\%$ of LLOQ)
 - Analyte Only (contribution $< 5\%$ of mean I.S.)
 - QC0 (contribution $< 20\%$ of LLOQ)
- Statistic prints:
 - Calibration curves
 - IS area Chart (Back to back injection)
 - Watson Std/QC from qualification run

Conclusions

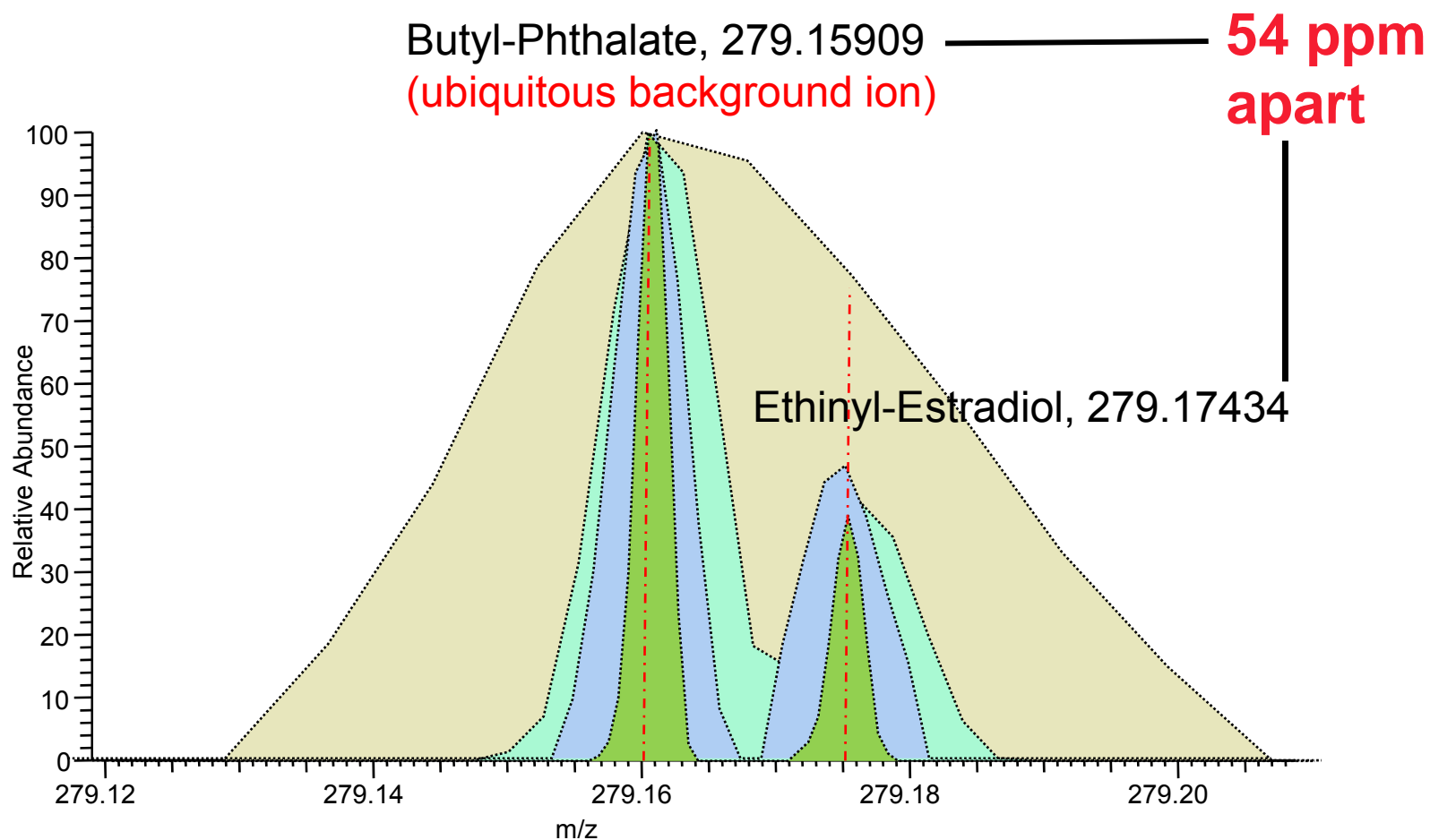
- Simple sample preparation and fast chromatography do not correlate to a robust and reliable bioanalytical method
- Protocol and screening driven MD approach allows rapid MD, less variability and more effective use of resources
- Phospholipids are important matrix effectors and monitoring them during MD is essential
- Proper documentation of all MD work is critical (for future MD, troubleshooting, auditing)

Next Generation Methods -HRMS

- Q Exactive – HRMS
 - Simpler method development
 - tSIM, Full Scan MS, SRM
 - Greater confidence in data - specificity
 - Troubleshooting for triple quadrupole based methods
 - Matrix effects
 - Metabolites
 - Co-eluting peaks
 - Qual Quan workflows
 - Metabolites
 - Metabolite stability
 - Biomarkers
 - Large and small molecule quantitation

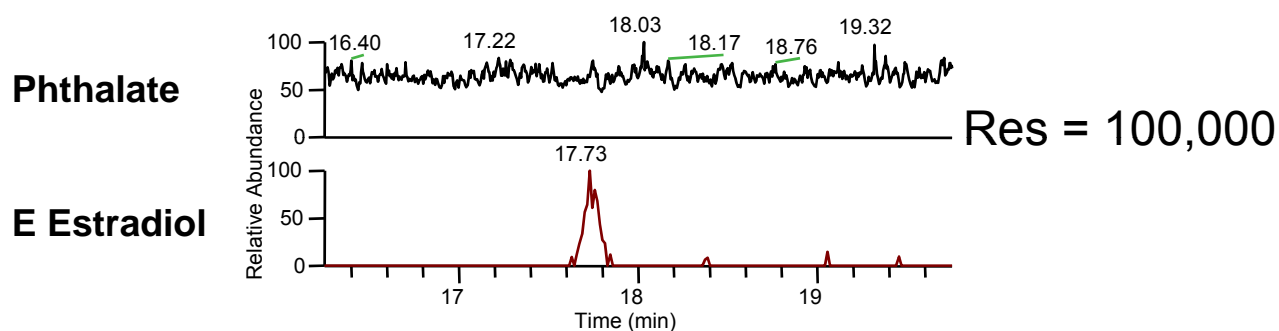
Specificity = Resolution + Mass Accuracy

Resolution: 10k, 30k, 50k, 100k

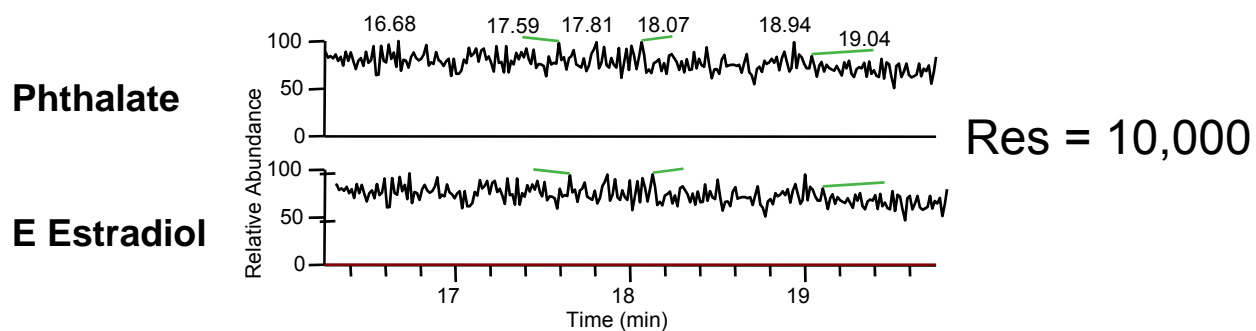
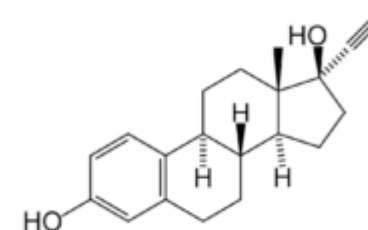


100 ppb Ethinyl-Estradiol – 100k vs 10 K Res

High Resolution and Mass Accuracy Essential



Ethinyl-Estradiol



Summary Table

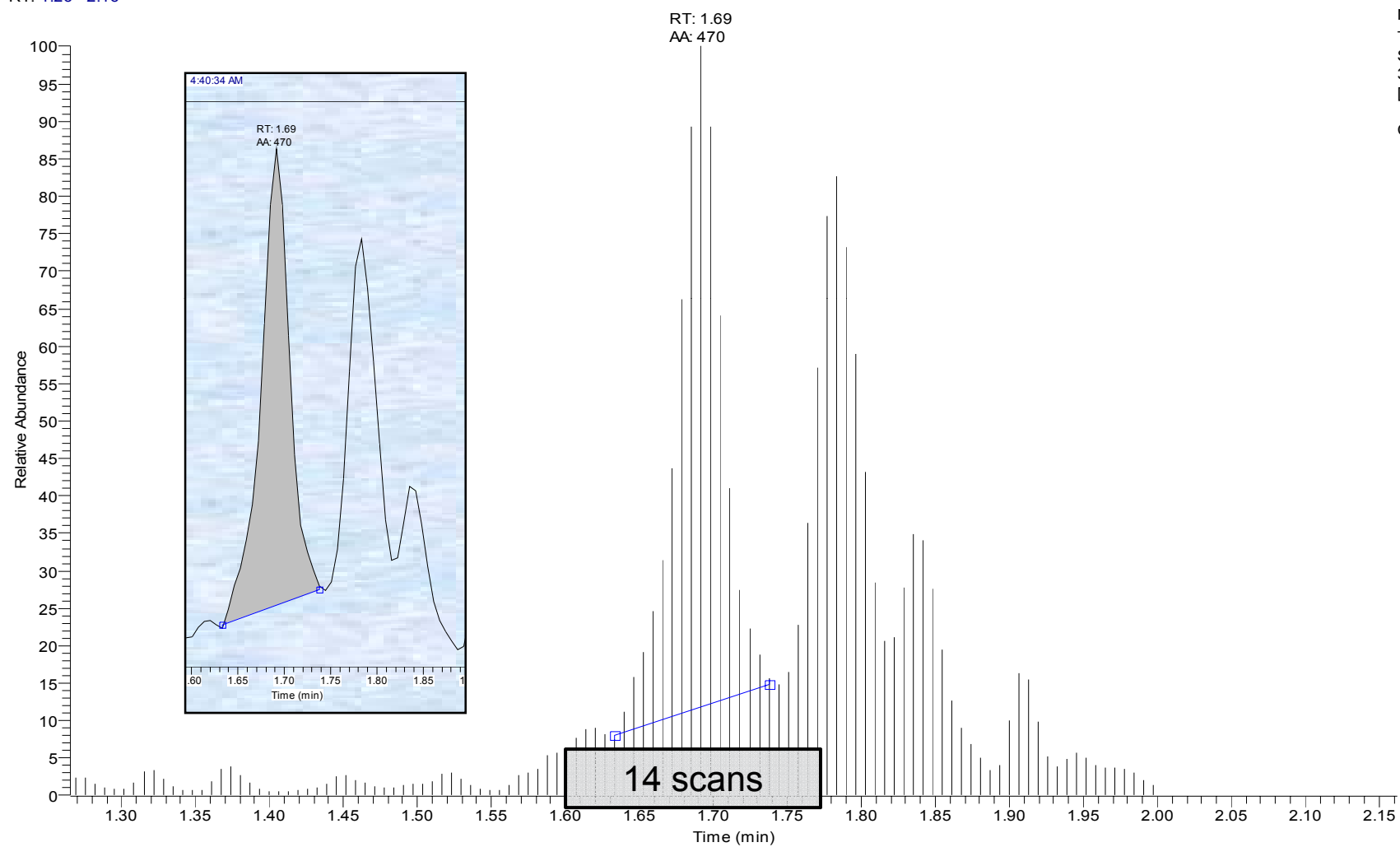
Compound	TSQ		QE Full Scan (70K)		QE tSIM (70K)	
	LOD	LLOQ	LOD	LLOQ	LOD	LLOQ
Oxycodone	1	5	5	10	1	5
Buprenorphine	50	100	50	50	50	50
Paroxetine	1	10	1	5	10	10
Ketoconazole	50	100	1	50	1	50
Clonazepam	1	10	5	50	10	10
Verapamil	1	1	5	5	1	5
Alprazolam	1	5	5	10	10	50
Reserpine	1	10	10	50	5	10
Clopidogrel	1	5	50	50	5	10

TSQ (SRM) Oxycodone LLOQ 5pg/mL

E:\VII Test\...\Rawfiles\CpdMix-Test-083
Std 5 pg/mL

8/5/2011 4:40:34 AM

RT: 1.26 - 2.16



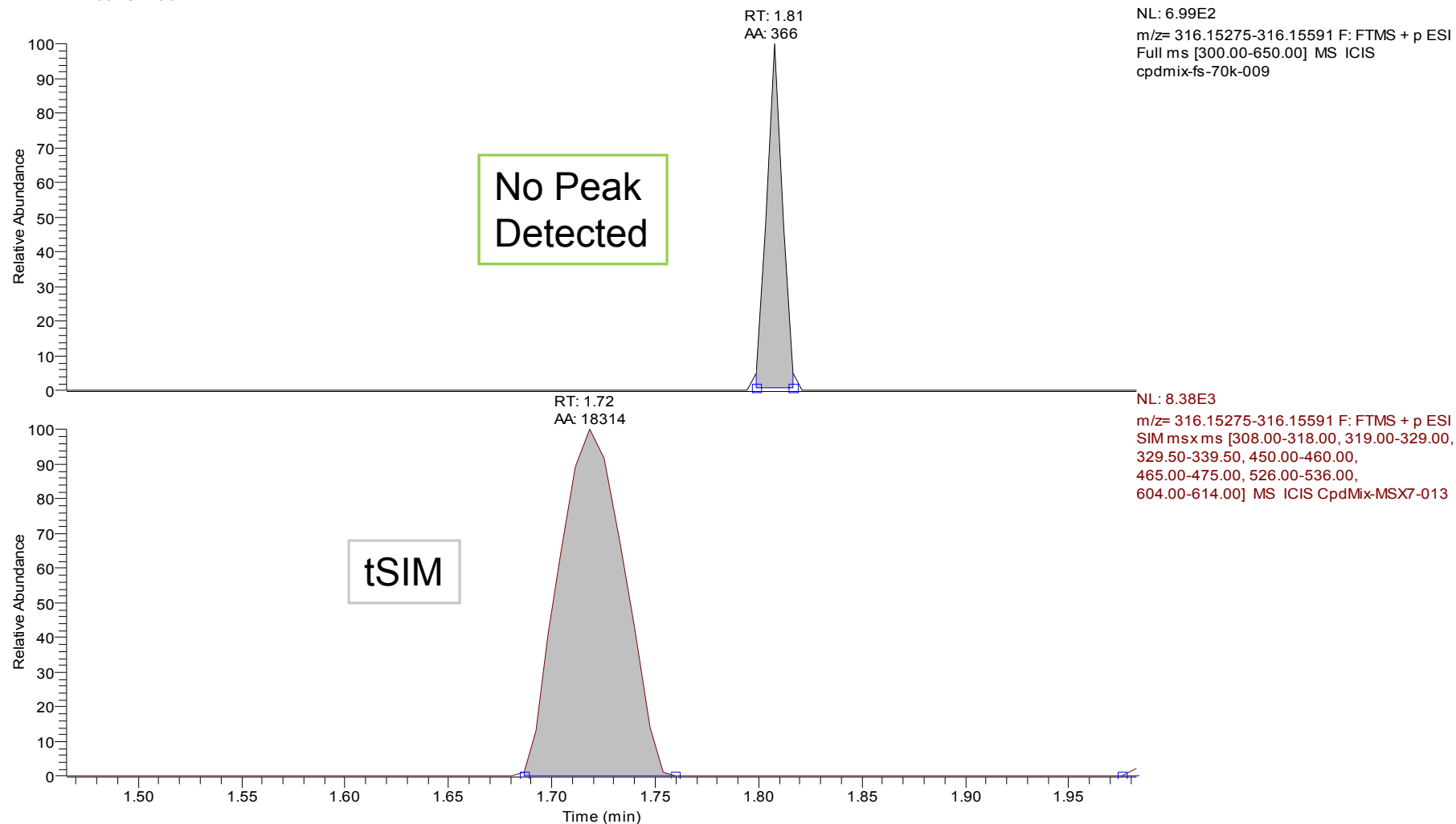
NL: 2.45E2
TIC F: + c ESI
SRM ms2
316.122
[241.083-241.085]
MS ICIS
CpdMix-Test-083

FS (5pg/mL) tSIM (5pg/mL) Comparison

cpdmix-fs-70k-009
Std 1 pg/mL

9/9/2011 2:24:33 AM

RT: 1.47 - 1.98 SM: 5G

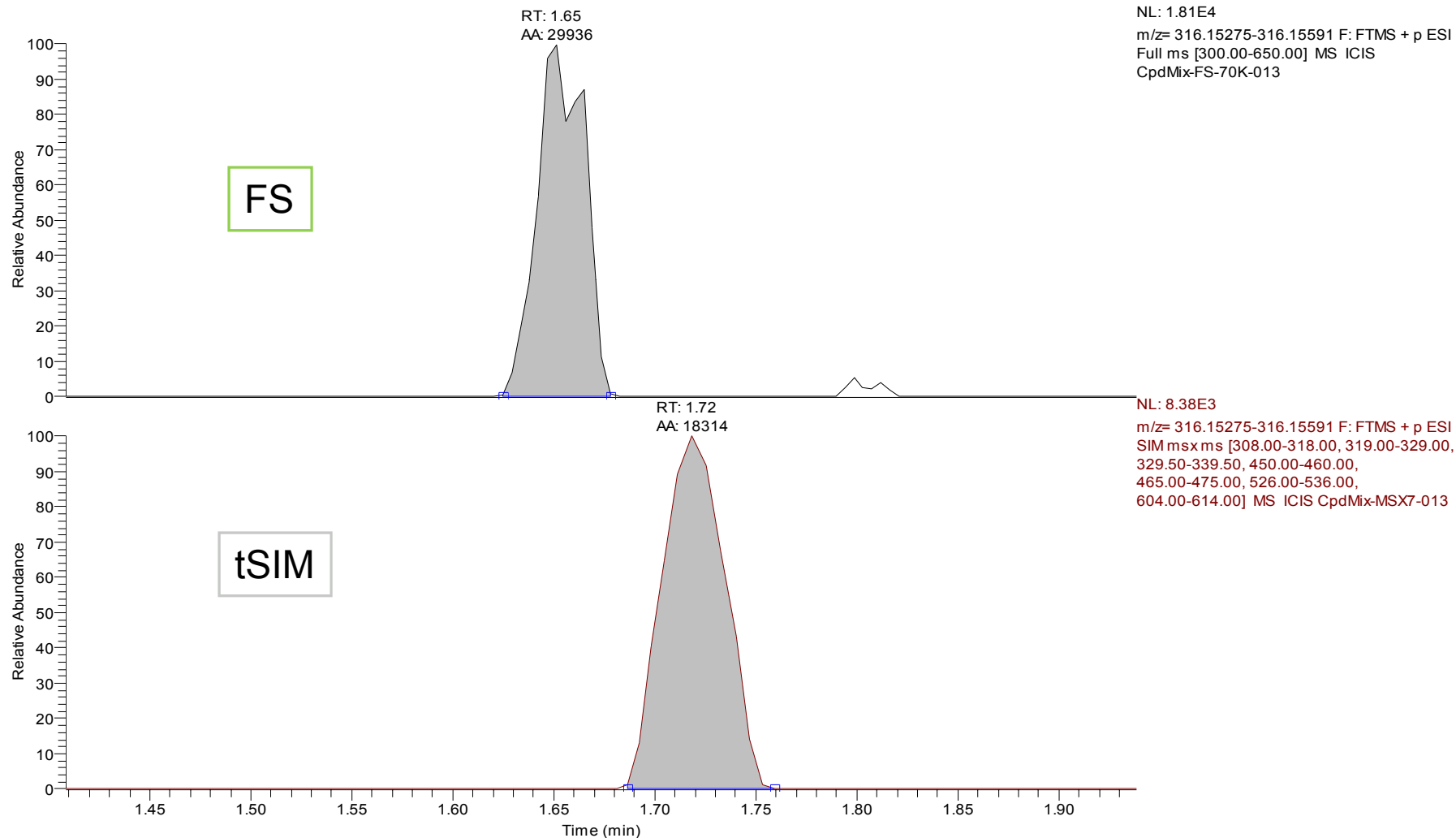


tSIM (5pg/mL) & FS (10pg/mL) Comparison

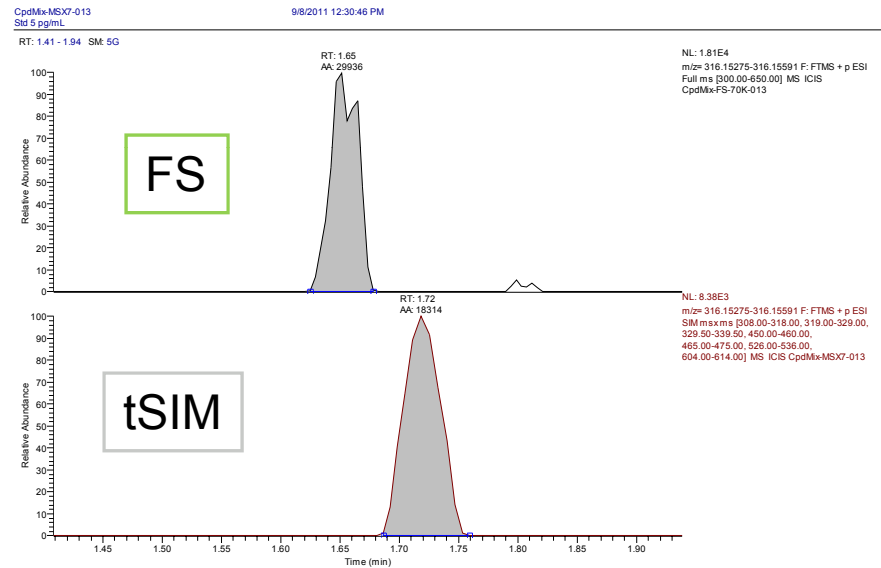
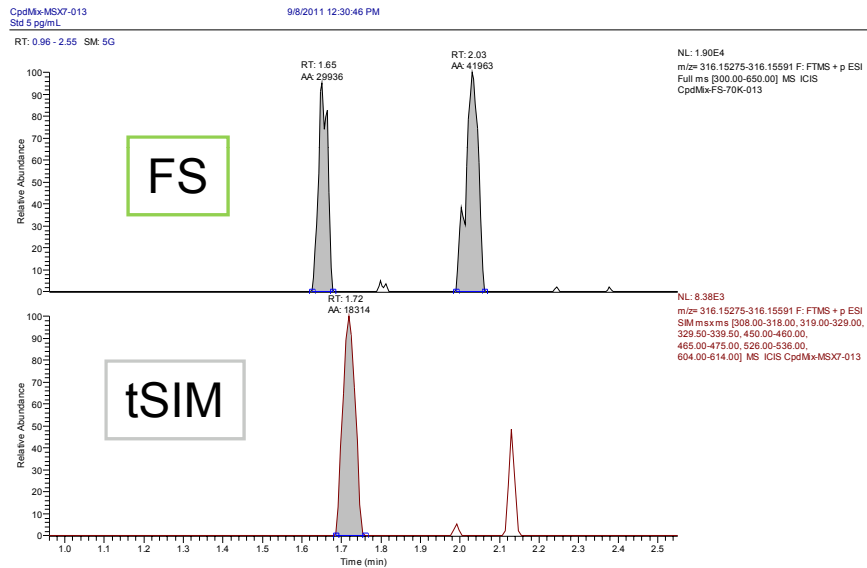
CpdMix-MSX7-013
Std 5 pg/mL

9/8/2011 12:30:46 PM

RT: 1.41 - 1.94 SM: 5G



tSIM (5pg/mL) & FS (10pg/mL) Comparison



Acknowledgement

- Min Meng, Ph.D. – Tandem Labs
- Scott Reuschel, M.F.S. – Tandem Labs
- KC Van Horne – Tandem Labs
- Kevin Cook – Thermo Fisher Scientific